

	Title: Cleaning and Disinfecting Stalls and Enclosures for Large/Agricultural Animals	SOP #	AV 50-104
		Version #	1
		Implementation Date	11/13/2016
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Standard Operating Procedure

1. Purpose

To establish minimum standards for cleaning and disinfecting stalls, runs and enclosures for animals in a large/agricultural animal setting.

2. Scope

All units providing animal care for large/agricultural animals must meet or exceed these minimum requirements for cleaning and disinfecting stalls, runs and enclosures based on the *Guide for the Care and use of Agricultural Animals in Research and Teaching*. Animals used in biomedical research and non-standard agricultural practice teaching must also meet the requirements in the Animal Welfare Act and the *Guide for the Care and Use of Laboratory Animals*.

3. Responsibilities

Principal Investigator and/or Facility Manager must develop standard operating procedures for cleaning and disinfecting animal stalls, runs, and enclosures and ensure compliance by animal care staff.

4. Procedure

Different levels of sanitation may be appropriate under different circumstances. Animals can harbor microbes that can be pathogenic to humans and other species. Hence, manure should be removed regularly unless a deep litter system or a built-up manure pack is being employed.

Animals housed in intensive laboratory environments should be kept clean and dry, and excreta should be removed on an appropriate schedule to achieve clean animals.

Stalls, runs and enclosures must be cleaned as needed (see husbandry SOPs for frequency), usually daily, to control pests, keep animals clean and dry, and maintain the air suitably free of dust and odors, especially ammonia.

Caution should be exercised with manure of animals infected with known pathogens; alternative methods of appropriate waste disposal should be used.

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Water delivery systems must be cleaned as needed to ensure adequate water supply and to prevent transmission of microbial or contaminant associated disease. Water delivery systems should be cleaned as needed to prevent algae or dirt from accumulating, preferably at least every 30 days.

It is important to monitor feeders daily to ensure that these are clean, free of moldy or wet feed, and not broken or damaged. The container should be cleaned out and soiled feed removed regularly.

Health and performance of animals can be affected by the time interval between successive occupations of intensive facilities. Complete disinfection of such quarters during the unoccupied phase of an all-in, all-out regimen of facility management is effective for disease management in some situations.

Where serious pathogens have been identified, the immediate environment may need to be disinfected as part of a preventive program. Elimination of moist and muddy areas in pastures may not be possible, but prolonged destocking is an available option. Drylot facilities may need to be scraped and refilled with uncontaminated materials. Thorough cleaning of animal housing facilities may be followed by disinfection. Selection of disinfection agents should be based on knowledge of potential pathogens and their susceptibilities to the respective agents.

When warranted, waste and bedding that have been removed from a site occupied by an animal that has died should be moved to an area that is inaccessible to other animals and the site appropriately disinfected.

For terminal cleaning, all organic debris should be removed from equipment and from floor, wall, and ceiling surfaces. If sanitation depends on heat for effectiveness, the cleaning equipment should be able to supply water that is at least 180°F. When chemical disinfection is used, the temperature of wash water may be cooler. If no machine is available, surfaces and equipment may be washed by hand with appropriate detergents and disinfectants and with vigorous scrubbing.

5. *References*

1. Institute of Laboratory Animal Resources: Guide for the Care and use of Laboratory Animals, National Academy Press, Eighth Edition (http://www.nap.edu/catalog.php?record_id=12910).



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2. Animal Welfare Act and Animal Welfare Regulations. United States Department of Agriculture
https://www.aphis.usda.gov/animal_welfare/downloads/Animal%20Care%20Blue%20Book%20-%202013%20-%20FINAL.pdf).
3. National Institutes of Health: Public Health Service Policy on Humane Care and Use of Laboratory Animals (<http://grants.nih.gov/grants/olaw/references/phspol.htm>).
4. The Federation of Animal Science Societies: Guide for the Care and Use of Agricultural Animals in Research and Teaching, 3rd Edition, 2010
http://www.fass.org/docs/agguide3rd/Ag_Guide_3rd_ed.pdf).

6. Definitions

1. AV – Attending Veterinarian – responsible for the health and well-being of all laboratory animals used at the institution
2. SOP – Standard Operating Procedure – established or prescribed methods to be followed routinely for the performance of designated operations or in designated situations
3. IACUC – Institutional Animal Care and Use Committee – responsible for assessment and oversight of the institution’s animal care program components and facilities
4. PI – Principal Investigator – a Central Oregon Community College employee having the background and training in scientific and administrative oversight necessary to conduct and manage the proposed study
5. Guide – the *Guide for the Care and use of Laboratory Animals*
6. AWA – Animal Welfare Act – requires that minimum standards of care and treatment be provided for certain animals bred for commercial sale, used in research, transported commercially, or exhibited to the public
7. PHS – Public Health Service – establishes guidelines for the proper care of animals to be used in research
8. ILAR – Institute for Laboratory Animal Research – provide the *Guide for the Care and Use of Laboratory Animals* to assist institutions in caring for and using animals in ways judged to be scientifically, technically, and humanely appropriate

Revision	Author	Revisions Made	Effective Date	Approval
1	Cindy Elston	• New SOP	11/13/16	Cindy Elston (AV)
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